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# Trivalent Copper Catalysis of the Autoxidation of Copper(II) Tetraglycine

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Oxygen reacts with copper(II) tetraglycine (G<sub>4</sub>) solutions at pH 7-9 to give  $Cu^{II}(H_{-3}G_4)^-$  (where  $H_{-3}$  refers to three deprotonated peptide nitrogens coordinated to copper) as an intermediate leading to the oxidation of the peptide. A dehydropeptide which hydrolyzes to diglycinamide and glyoxylglycine is the major product of the peptide oxidation. Copper(III) tetraglycine is the catalyst for the autoxidation, and the rate of decomposition of  $Cu^{III}(H_{-3}G_4)^-$  controls the rate of  $O_2$  uptake. This process leads to an autocatalytic increase in Cu(III) concentration. In the proposed mechanism the autoxidation reactions are initiated by traces (<10<sup>-7</sup> M) of  $Cu^{III}(H_{-3}G_4)^-$  which form by the disproportionation of  $Cu^{II}(H_{-2}G_4)^-$  into Cu(II) and Cu(III) complexes. It is proposed that the Cu(III) complex decomposes to give a species which reacts rapidly with  $O_2$  and eventually generates more Cu(III) complex as well as the oxidized peptide. Copper is an effective catalyst in this autoxidation because of its multiple oxidation states and because Cu(III) complexes serve as a reservoir of oxidizing power.

## Introduction

Copper(II) and tetraglycine  $(G_4^-)$  form complexes with a stepwise loss of peptide hydrogen ions occurring at  $pK_a$  values of 5.52, 6.78, and 9.16 to give  $Cu^{II}(H_{-1}G_4)$ ,  $Cu^{II}(H_{-2}G_4)^-$ , and  $Cu^{II}(H_{-3}G_4)^{2-}$ , respectively.<sup>1</sup> The crystal structure of Na<sub>2</sub>Cu<sup>II</sup>(H\_{-3}G\_4)·10H<sub>2</sub>O shows that the amine nitrogen and three deprotonated-peptide nitrogens are bound to copper in a nearly square-planar arrangement and that the carboxylate group is not coordinated.<sup>2</sup> The Cu<sup>II</sup>(H\_{-3}G\_4)^{2-} complex is relatively labile, reacting very rapidly with acids and somewhat more slowly with solvent and nucleophiles.<sup>3</sup> The triply deprotonated form is easily oxidized to Cu<sup>III</sup>(H\_{-3}G\_4)^- using IrCl<sub>6</sub><sup>2-</sup> or electrochemical techniques.<sup>4,5</sup> Its electrode potential (Cu<sup>III</sup> + e<sup>-</sup> → Cu<sup>II</sup>) is 0.63 V (vs. NHE). The Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> complex is slow to undergo in-plane substitution reactions but is very rapid in its electron-transfer reactions.<sup>6</sup>

Copper(II) ion activates the reaction of molecular oxygen with tetraglycine wherein the peptide is oxidized to diglycinamide ( $G_{2a}$ ) and glyoxylglycine.<sup>7</sup> The reaction with  $O_2$ is spontaneous in neutral solution and is autocatalytic. Copper(III)-peptide complexes are observed as intermediates during the autoxidation. Nickel(II) tetraglycine undergoes similar reactions with  $O_2$ ,<sup>8</sup> forming Ni<sup>III</sup>( $H_{-3}G_4$ )<sup>-</sup> as an intermediate,<sup>9</sup> but with the nickel complex the  $G_4$  is oxidized to triglycinamide, carbon dioxide, and formaldehyde. In the present work copper(III) peptide intermediates are shown to be the catalysts for the autoxidation. The rate of  $O_2$  uptake is controlled by the rate of decomposition of  $Cu^{III}(H_{-3}G_4)^{-}$ . Subsequent reactions lead to the formation of more  $Cu^{III}$ .  $(H_{-3}G_4)^{-}$ .

There have been a number of studies of the autoxidation of cobalt(II) di- and tripeptide complexes.<sup>10-17</sup> In these systems oxygenation produces  $\mu$ -peroxo binuclear Co(III) complexes. Although this oxygenation can be reversible, the binuclear complexes decompose irreversibly to mononuclear Co(III) species.<sup>13,16</sup> The oxidation of ascorbic acid by oxygen in the cobalt(II)-diglycine system<sup>18</sup> and of glycyltryptophan by oxygen in the cobalt(II)-glycyltryptophan system<sup>11</sup> has been described as catalytic. In the present system the copper(III) peptide serves as a pool of reactive oxidizing power and generates an intermediate which is very reactive with  $O_2$ . This intermediate could be a carbon-centered free radical ( $Cu^{II}R$ .) which reacts with  $O_2$  to form a peroxy radical (Cu<sup>II</sup>RO<sub>2</sub>). Similar R. and  $RO_2$  species (but without metal ions) are proposed as chain centers in the autoxidation of N-alkylamides to give hydroperoxides.<sup>19-21</sup> The N-alkylamide reactions are slow and require elevated temperatures. The copper-catalyzed autoxidation of tetraglycine in the present system is much more rapid because Cu(III) assists the formation of  $Cu^{II}R$ .

An alternative mechanism is possible in which the copper(III) peptide generates a copper(I) dehydropeptide as an intermediate which reacts with  $O_2$  to form a copper(III) peroxide. In this mechanism the reactive intermediates are copper(I) and copper(III) peroxide rather than radicals centered at the  $\alpha$ -carbon atom. The reaction products are the same and the detailed rate expressions fit both mechanisms.

It is interesting to note that copper(II) is also known to catalyze the oxidative degradation of protein and poly- $\alpha$ -amino acids by H<sub>2</sub>O<sub>2</sub><sup>22</sup> and the autoxidation of petroleum products.<sup>23</sup> There are also a number of copper-containing enzymes whose function, so far as known, involves catalysis of O<sub>2</sub> oxidation of various substrates.<sup>24</sup>

### **Experimental Section**

**Reagents.** Tetraglycine was obtained from Biosynthetika (Oberdorf, Switzerland). Copper(II) perchlorate was prepared from reagent grade copper carbonate and perchloric acid. Stock solutions were standardized by EDTA/murexide titration. Solutions of copper(II) tetraglycine were prepared with a 5–10% weight excess of ligand to suppress precipitation of copper hydroxide.

An electrochemical flow cell, described previously,<sup>25</sup> was used to generate  $Cu^{III}(H_{-3}G_4)^-$  for the  $Cu^{III}$  decomposition and oxygen-uptake studies. A solution of  $Cu^{II}(H_{-3}G_4)^2^-$  initially at pH 10.4–10.6 in 0.10 M NaClO<sub>4</sub> was passed through the working electrode (maintained at +0.66 V vs. Ag/AgCl) at 0.6–1.2 mL/min. The pH of the eluent was between 6 and 8.

**Kinetic Experiments.** Most kinetic measurements were thermostated at  $25.0 \pm 0.1$  °C. A few reactions were carried out at 37 °C. Ionic strength was maintained essentially constant with 0.10 M NaClO<sub>4</sub> supporting electrolyte. All pH measurements were performed on a Radiometer Model 26 pH meter. Oxygen uptake was followed with a Yellow Springs Model 53 polarographic O<sub>2</sub> monitor.

The formation and decomposition of Cu(III) were followed using a Cary Model 16 spectrophotometer. The molar absorptivity of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> was found to be 7400 M<sup>-1</sup> cm<sup>-1</sup> at 365 nm. For the initiation experiments, solutions of Cu<sup>II</sup>G<sub>4</sub> were prepared at pH 8.60 in 0.010 M H<sub>3</sub>BO<sub>3</sub> buffer (0.1 M NaClO<sub>4</sub>) and allowed to equilibrate for 2–12 h under nitrogen. Solutions were then passed through 100-mµ Millipore filters and again allowed to equilibrate under N<sub>2</sub> for about 12 h. Reactions were initiated by flowing solutions through a double two-jet tangential mixer into a 5.00-cm observation cell. The initial absorbance was measured by mixing the Cu<sup>II</sup>G<sub>4</sub> solution with a N<sub>2</sub>-saturated 0.10 M NaClO<sub>4</sub> solution. For the autoxidation studies Cu<sup>II</sup>G<sub>4</sub> solutions were mixed with O<sub>2</sub>-saturated 0.10 M NaClO<sub>4</sub> solutions.

Chromatographic Peptide Analysis. The products of ligand oxidation were determined chromatographically on a Beckman Model 120B amino acid analyzer. Samples were passed down a 15-cm column of Beckman PA 35 or Hamilton H70 cation-exchange resin and eluted with pH 3.2 citrate buffer ( $0.2 \text{ M Na}^+$ , 0.067 M citrate). The instrument was calibrated as to elution time and detector sensitivity with standard solutions of glycine and di-, tri-, and tetraglycine as well as peptide amide fragments of tetraglycine. This method detects only those oxidation products which contain amine nitrogens.

Analysis for Carbonyl. The autoxidation products were analyzed for carbonyl content by the method of Anbar, Levitzki, and Berger.<sup>26</sup> A 1-mL sample was mixed with 10 mL of 5% trichloroacetic acid



**Figure 1.** Progress of autoxidation of copper(II) tetraglycine at 25 °C is monitored by  $O_2$  uptake and Cu(III) formation and decay. A solution of  $2.5 \times 10^{-3}$  M [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub> at pH 7.50 in 0.018 M 2,6-lutidine buffer is initially saturated with  $O_2$  (1 atm) and the sample is divided. The  $O_2$  concentration is followed polarographically and the concentration of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> is monitored from the absorbance at 365 nm in a 1-cm cell.

Table I. Effect of Copper(III) Tetraglycine on the Induction Time for  $O_2$  Uptake by Copper(II) Tetraglycine

[Cu <sup>III</sup> - (H <sub>-3</sub> G <sub>4</sub> ) <sup>-</sup> ], <sup>a</sup> M	tind, <sup>b</sup> min	[Cu <sup>III</sup> - (H <sub>-3</sub> G <sub>4</sub> ) <sup>-</sup> ], <sup><i>a</i></sup> M	tind, <sup>b</sup> min	
0	187	$2.5 \times 10^{-4}$	12	
$2.5 \times 10^{-5}$	48	$5.0 \times 10^{-4}$	8	
$1.25 \times 10^{-4}$	18			

<sup>a</sup> Electrochemically prepared  $Cu^{III}(H_{-3}G_4)^-$  added to 5.0 × 10<sup>-3</sup> M [ $Cu^{II}G_4$ ]<sub>T</sub> at pH 8.2. <sup>b</sup> Time required to consume 5% saturation of  $O_2$  at 25.0 °C.

and 1 mL of 0.1% 2,4-dinitrophenylhydrazine in 2 M HCl. After 15 min, 20 mL of ethyl acetate was added, and the phases were mixed for 20 s. Then 3.0 mL of 10%  $Na_2CO_3$  was added and the phases were again mixed. After phase separation, 5.0 mL of the aqueous phase was added to 4.0 mL of 2 M NaOH, and absorbance was measured at 440 nm after an additional 15 min.

#### Results

Initiation and Inhibition of Autoxidation. When a solution of copper(II) tetraglycine, pH 7-9, is saturated with  $O_2$ , there is little  $O_2$  uptake observed initially. However, after an induction period (Figure 1) the  $O_2$  uptake rate increases autocatalytically until the depletion of  $O_2$  causes the rate to decrease again. Below pH 7 and above pH 10 no measurable amounts of  $O_2$  are consumed over a period of 3-4 h at 25 °C.

The induction time (which we will define as the time required for the O<sub>2</sub> concentration to decrease by 5%) becomes smaller as the concentration of  $[Cu^{II}G_4]_T$  increases, where  $[Cu^{II}G_4]_T = [Cu^{II}(H_{-3}G_4)^{2-}] + [Cu^{II}(H_{-2}G_4)^{-}] + [Cu^{II}(H_{-1}G_4)]$ , and as the temperature increases. The induction time is shortened by the addition of small quantities of oxidants such as  $H_2O_2$  and  $K_2S_2O_8$ . Small amounts of electrochemically prepared  $Cu^{III}(H_{-3}G_4)^{-}$  greatly shorten the induction time (Table I), and larger amounts cause the maximum rate of  $O_2$  uptake to be established immediately.

As reported previously,<sup>7</sup> room light inhibits the reaction, greatly lengthening the induction time for  $O_2$  uptake. Hence the present reactions are carried out in the dark. However, once autoxidation becomes rapid, the system becomes relatively insensitive to room light or even to brief exposures of intense light. Longer exposure (~500 s) to a 75-W tungsten iodide lamp causes photodecomposition of  $Cu^{III}(H_{-3}G_4)^-$  and its loss quenches the  $O_2$  reaction until another induction period elapses. [The  $O_2$  monitor is very sensitive to temperature, and intense light can cause a spurious response even with a thermostated compartment. This effect probably caused what appeared to be a rapid on/off photoinhibition reported earlier.<sup>7</sup> Additional photochemical studies are needed, but it is clear that Cu(III) species are photochemically active.]

The O<sub>2</sub> uptake reaction is stopped by the addition of reducing agents (SO<sub>3</sub><sup>2-</sup>, ascorbic acid, hydroquinone, I<sup>-</sup>, SCN<sup>-</sup>, or di-tert-butyl nitroxide). However, Br- has little effect. DPPH (1,1-diphenyl-2-picrylhydrazyl) is a stable nitrogen free radical only slightly soluble in water and is frequently used as a radical scavenger. The effect of trace concentrations of DPPH  $(2 \times 10^{-7}-1.5 \times 10^{-6} \text{ M})$  on the reaction of  $[\text{Cu}^{11}\text{G}_4]_T$ with  $O_2$  is studied at 37 °C. The length of the induction time is in direct proportion to the amount of DPPH added. However, after the induction period, the maximum rate of  $O_2$ uptake is not altered by the DPPH. The induction period at various levels of DPPH is independent of the O<sub>2</sub> concentration and is independent of pH between 7.5 and 9.0. However, the induction period is shortened as the  $[Cu^{11}G_4]_T$  concentration is increased. When  $[Cu^{11}G_4]_T$  is  $3.4 \times 10^{-3}$  M, pH 8.0, 37 °C, the rate of loss of DPPH (measured from the difference in induction periods) is  $1.1 \times 10^{-9}$  M s<sup>-1</sup>.

The autoxidation also can be followed by measuring the absorbance increase at 365 nm, which is due to the formation of Cu(III) and is observable within a few minutes after mixing  $[Cu^{11}G_4]_T$  with O<sub>2</sub>. This exponential absorbance increase is a more sensitive measure of the progress of the autoxidation than is O<sub>2</sub> uptake. During the apparent induction period observed with the O<sub>2</sub> monitor, there is a continuous increase in the concentration of Cu(III) species.

Identity of the Reactants and Intermediates. For the pH range over which the autoxidation occurs the main copper(II) tetraglycine species is  $Cu^{II}(H_{-2}G_4)^-$  with smaller concentrations of  $Cu^{II}(H_{-3}G_4)^{2^-}$  and  $Cu^{II}(H_{-1}G_4)$  present. These species rapidly attain equilibrium.<sup>3</sup> The possibility of an O<sub>2</sub> complex of  $Cu^{II}(H_{-2}G_4)^-$  was considered, but we could not find evidence for measurable amounts of such a species. Two methods of detection were used. Difference spectra for N<sub>2</sub>-saturated vs. O<sub>2</sub>-saturated solutions of  $Cu^{II}(H_{-2}G_4)^-$  failed to show absorbance changes which might be assigned to an O<sub>2</sub> complex, although the slow growth of a Cu(III) complex was detected. The O<sub>2</sub> monitor failed to show any immediate O<sub>2</sub> uptake and failed to show any O<sub>2</sub> release upon acidification of freshly mixed solutions of  $Cu^{II}(H_{-2}G_4)^-$  and O<sub>2</sub>. The latter results indicate that if a complex of  $[Cu(H_{-2}G_4) \cdot O_2]^-$  exists, it has a stability constant less than 10 M<sup>-1</sup> (based on the sensitivity of the O<sub>2</sub> monitor).

Appreciable concentrations of a Cu(III) complex are formed slowly when solutions of Cu<sup>II</sup>( $H_{-2}G_4$ )<sup>-</sup> (10<sup>-3</sup> M or higher) are saturated with O<sub>2</sub>. Evidence for the existence of trivalent copper complexes and a description of their properties are given in other work.<sup>4,5,7</sup> The complex formed in this autoxidation process is identified as Cu<sup>III</sup>( $H_{-3}G_4$ )<sup>-</sup> based on its redox properties, its spectrum, and the kinetics of its decomposition. The Cu(III) species formed by the O<sub>2</sub> reactions decomposes at the same rate, with the same pH dependence, and the same Cu<sup>II</sup>( $H_{-2}G_4$ )<sup>-</sup> dependence (described later) as Cu<sup>III</sup>( $H_{-3}G_4$ )<sup>-</sup> solutions prepared electrochemically.

After the completion of the oxygenation and the decay of the  $Cu^{III}(H_{-3}G_4)^-$  complex, a new species is observed in the peptide analyzer. The elution time for this species is similar to that for  $G_4$ , but after mild hydrolysis it is converted mainly to glycylglycinamide (GGa) and to an equivalent amount of a carbonyl compound. The intermediate is believed to be a dehydropeptide (G<sub>4</sub>DHP) which hydrolyzes according to eq

 $NH_2CH_2CONHCH_2CON = CHCONHCH_2COO^{-} \rightarrow NH_2CH_2CONHCH_2CONH_2 + HCOCONHCH_2COO^{-}$ (1)

1 to give GGa and glyoxylglycine. A species with the same elution time, which also hydrolyzes to give GGa and carbonyl, is the sole oxidized ligand product found in the acid decomposition of  $Cu^{III}(H_{-3}G_4)^{-,27}$  We believe this intermediate

Table II.	Stoichiometry	of Autoxidation	of
Copper (II	) Tetraglycine <sup><math>a</math></sup>		

copper(ii) Tettagiyene						
amt O	of a	mtof a G₄	umt of G2a			
consu × 10	imed con <sup>4</sup> , M ×	nsumed pr 10⁴, M ×	oduced $\Delta$ 10 <sup>4</sup> , M $\Delta$	$\Delta[G_4]/\Delta[O_2]$	[G <sub>2</sub> a]/ [O <sub>2</sub> ]	
2.4 4.8 7.3	45 33 38	5.90 9.20 13.8	3.25 5.88 9.20	2.4 1.9 1.9	1.3 1.2 1.3	

 $^a$  2.5  $\times$  10<sup>-3</sup> M  $[Cu^{II}G_a]_T$  at pH 8.6 allowed to take up O<sub>2</sub> from saturated solution; reaction quenched by sweeping with N<sub>2</sub>.

to be the same in both cases. A similar type of reaction was observed in the decomposition of  $Ni^{III}(H_{-3}G_4)^-$  except that the oxidation occurred at a carboxylate terminal giving as the initial products CO<sub>2</sub> and a *N*-(hydroxymethyl)amide, H<sub>2</sub>NCH<sub>2</sub>CONHCH<sub>2</sub>CONHCH<sub>2</sub>CONHCH<sub>2</sub>CONHCH<sub>2</sub>OH, which hydrolyzed to G<sub>3</sub>a and HCHO.<sup>9</sup> Neither CO<sub>2</sub> nor HCHO was found in the Cu-G<sub>4</sub> oxidation products.

**Products and Stoichiometry of the Autoxidation.** After the oxygenation is complete (and the  $Cu^{III}(H_{-3}G_4)^-$  species is decomposed), the solutions no longer have oxidizing ability. Tests for the presence of hydrogen peroxide among the oxidation products were negative. The fluorometric test used<sup>28</sup> is capable of detecting hydrogen peroxide at extremely low levels. Therefore all the oxygen consumed during the reaction is reduced to water and the oxidized ligand products.

The ultimate products of the autoxidation after acidification are Cu(II), GGa, and glyoxylglycine. Smaller quantities of other fragments including glycylglycine and glycinamide are found. These are the same products observed at pH 7–9 for the decomposition (in the absence of  $O_2$ ) of electrochemically prepared Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-,27</sup> The observation that oxidation of the ligand occurs predominantly at the third peptide residue from the amine terminal is also consistent with results using IrCl<sub>6</sub><sup>2-</sup> oxidation of copper(II) peptide complexes.<sup>26,29</sup>

The stoichiometry of the reaction was determined by measuring the  $O_2$  uptake and then quenching the reaction by sweeping the solution with  $N_2$ , when 20, 40, and 60% of the  $O_2$  initially present had been consumed. These samples were analyzed for their carbonyl products and for their ninhydrin-active peptide products. The results in Table II show that the ratio of  $G_4$  lost to  $O_2$  consumed is approximately 2:1 and that GGa accounts for 55–67% of the  $G_4$  consumed. Most of the remaining ninhydrin activity is recovered as  $G_2$ , G, and Ga, although a small decrease from the initial level of ninhydrin-active material is observed. The overall reaction stoichiometry observed is expressed in eq 2 where x represents

 $2G_4 + O_2 \rightarrow$ 

$$1.3GGa + 1.3(HCOCONHCH_{2}COO^{-}) + x$$
 (2)

a mixture of other products resulting from the oxidation of  $G_4$ .

**Kinetic Measurements.** The processes of Cu(III) formation, Cu(III) decomposition, and  $O_2$  consumption all occur simultaneously during the autoxidation. In analyzing the kinetics of the system we have, insofar as possible, studied these processes independently.

**Decomposition of Cu<sup>III</sup>(H**<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> in the Absence of O<sub>2</sub>. The decomposition of electrochemically generated Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> in the absence of O<sub>2</sub> between pH 7 and 9 gives a recovery of approximately 50% G<sub>4</sub> and gives 33% GGa and other oxidized ligand products as in eq 2. The loss of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> is first order in Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> when the concentration of copper(II) tetraglycine is held constant. Since copper(II) tetraglycine is one of the products of the reaction (at pH 7–9), its concentration may be maintained essentially constant either by carrying out the reaction in the presence of a large excess of copper(II) tetraglycine or by scavenging the copper(II) tet-



**Figure 2.** Dependence of the first-order rate constants  $(k_{obsd})$  for  $Cu^{III}(H_{-3}G_4)^-$  formation (in the presence of  $O_2$ ) and for  $Cu^{III}(H_{-3}G_4)^-$  decomposition (in the absence of  $O_2$ ) on the concentration of  $[Cu^{II}G_4]_T$ . Conditions: pH 8.60 in 0.01 M  $[H_3BO_3]_T$ , 0.10 M NaClO<sub>4</sub>, 25.0 °C; formation at 1.2 × 10<sup>-3</sup> M O<sub>2</sub>; decomposition with initial  $[Cu^{III}_{-4}(H_{-3}G_4)^-]$  of 1.0 × 10<sup>-5</sup> M and no  $O_2$ .



Figure 3. Autocatalytic formation of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> where the reaction is first order in product.  $A_0$  is the initial absorbance (365 nm) and  $A_t$  is the absorbance at time t after mixing with dissolved O<sub>2</sub>. Conditions: pH 8.60 in 0.01 M [H<sub>3</sub>BO<sub>3</sub>]<sub>T</sub>, 0.10 M NaClO<sub>4</sub>, 25.0 °C,  $5 \times 10^{-4}$  M [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub>,  $6 \times 10^{-4}$  M O<sub>2</sub>.

raglycine as it forms. Data for the decomposition of  $1 \times 10^{-5}$  M Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> in the presence of copper(II) tetraglycine are plotted in Figure 2. As the [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub> concentration increases above  $10^{-4}$  M its effect reaches a limiting value and the observed rate constant for Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> decomposition becomes independent of the [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub>. The intercept in Figure 2 was measured in the presence of  $10^{-4}$  M EDTA as a scavenger for [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub> produced during the decomposition. At this concentration EDTA does not affect the decomposition rate itself and the Cu<sup>II</sup>EDTA<sup>2-</sup> formed does not catalyze the reaction. Hence excellent first-order plots are obtained with a rate constant of  $4 \times 10^{-5}$  s<sup>-1</sup> in borate buffer at pH 8.6.

Kinetics of Cu<sup>III</sup>( $\mathbf{H}_{-3}\mathbf{G}_{4}$ ) Formation in the Autoxidation. The formation of Cu<sup>III</sup>( $\mathbf{H}_{-3}\mathbf{G}_{4}$ )<sup>-</sup> was observed by following the 365-nm absorbance ( $\mathcal{A}_{i}$ ) of an O<sub>2</sub> saturated solution of copper(II) tetraglycine undergoing autoxidation. When these data were plotted as ln ( $\mathcal{A}_{i} - \mathcal{A}_{0}$ ) vs. time, where  $\mathcal{A}_{0}$  is the absorbance before O<sub>2</sub> is added, plots were obtained with long linear segments as shown in Figure 3. This plot corresponds to the rate expression (3) where  $k_{obsd}$  is a first-order rate

$$d[Cu^{III}(H_{-3}G_4)^{-}]/dt = k_{obsd}[Cu^{III}(H_{-3}G_4)^{-}]$$
(3)

constant but the reaction is first order in product and hence is autocatalytic. Early in the reaction the difference between  $A_t$  and  $A_0$  is very small and the same type of scatter is observed as found near the end of first-order reactions (where  $A_t$  and  $A_{\infty}$  approach each other). The integrated rate expression derived from eq 3 requires some finite concentration of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> at the beginning of the reaction. The intercept

Table III. Rate Constants for Autocatalytic Cu<sup>III</sup>( $H_{-3}G_4$ )<sup>-</sup> Formation at 25.0 °C and 0.10 M NaClO<sub>4</sub>

run	pН	buffer medium	[Cu <sup>II</sup> G₄] <sub>T</sub> , M	[O <sub>2</sub> ] <sub>0</sub> , M	kobsd, s <sup>-1</sup>
1	7.0	unbuffered	$5.0 \times 10^{-4}$	$1.2 \times 10^{-3}$	5.5 × 10 <sup>-4</sup>
2	8.0	unbuffered	2.0 × 10⁻³	$1.2 \times 10^{-3}$	5.6 × 10 <sup>-4</sup>
3	8.0	unbuffered	$2.0 \times 10^{-4}$	$1.2 \times 10^{-3}$	5.1 × 10 <sup>-4</sup>
4	8.6	borate	$8.2 \times 10^{-4}$	$1.2 \times 10^{-3}$	5.5 × 10⁻⁴
5	8.6	borate	4.1 × 10⁻⁴	$1.2 \times 10^{-3}$	5.1 × 10 <sup>-4</sup>
6	8.6	borate	$2.1 \times 10^{-4}$	$1.2 \times 10^{-3}$	$4.7 \times 10^{-4}$
7	8.6	borate	1.6 × 10 <sup>-4</sup>	$1.2 \times 10^{-3}$	$4.2 \times 10^{-4}$
8	8.6	borate	$1.0 \times 10^{-4}$	$1.2 \times 10^{-3}$	3.6 × 10⁻⁴
9	8.6	borate	8.2 × 10 <sup>-5</sup>	$1.2 \times 10^{-3}$	$3.0 \times 10^{-4}$
10	8.6	borate	4.1 × 10 <sup>-5</sup>	$1.2 \times 10^{-3}$	1.4 × 10⁻⁴
11	8.6	borate	2.1 × 10⁻⁴	$1.2 \times 10^{-3}$	4.5 × 10⁻⁴
12	8.6	borate	2.1 × 10⁻⁴	6.0 × 10 <sup>-4</sup>	4.1 × 10⁻⁴
13	8.6	borate	2.1 × 10 <sup>-4</sup>	2.4 × 10⁻⁴	$3.3 \times 10^{-4}$
14	8.6	borate	$2.1 \times 10^{-4}$	$1.2 \times 10^{-4}$	$2.5 \times 10^{-4}$



Figure 4. Dependence of the initial rate of O<sub>2</sub> uptake on [Cu<sup>III</sup>-(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup>]. Solid line calculated from eq 7 with  $k_1 = 3.6 \times 10^{-4} \text{ s}^{-1}$  and  $k_3/k_2 = 0.5$ . Conditions: pH 7.5 in 0.018 M 2,6-lutidine buffer, 0.10 M NaClO<sub>4</sub>, 25.0 °C, 2.5 × 10<sup>-3</sup> M [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub>, 1.1 × 10<sup>-3</sup> M [O<sub>2</sub>]<sub>t</sub>.

in Figure 3 corresponds to a value of  $5 \times 10^{-8}$  M Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup>.

Values for the autocatalytic rate constant  $(k_{obsd})$  under a variety of conditions are given in Table III. The first four values show that  $k_{obsd}$  does not change significantly with pH, the presence or absence of borate buffer, and the initial  $[Cu^{II}G_4]_T$  concentration, as long as Cu(II) is greater than 2 × 10<sup>-4</sup> M. Runs 4–10 show that as the  $[Cu^{II}G_4]_T$  concentration is reduced below 2 × 10<sup>-4</sup> M, the  $k_{obsd}$  values decrease (see also Figure 2). Runs 11–14 show that  $k_{obsd}$  decreases somewhat as the initial O<sub>2</sub> concentration is lowered.

Kinetics of  $O_2$  Uptake. Studying the kinetics of oxygen uptake during autoxidation of  $Cu^{II}G_4$  posed difficulties because several hours may elapse before appreciable rates of O2 uptake are observed. The addition of traces of electrochemically prepared  $Cu^{III}(H_{-3}G_4)^-$  causes  $O_2$  consumption to proceed at measurable rates without an induction period. It was thus possible to circumvent the irreproducibility of working with a system of uncertain composition by measuring the initial rate of  $O_2$  uptake of synthetic mixtures of  $Cu^{II}G_4$  and electro-chemically prepared  $Cu^{III}(H_{-3}G_4)^-$ . It is observed that at higher  $[Cu^{II}G_4]_T$  concentrations the rate of O<sub>2</sub> uptake is essentially independent of Cu(II) concentration, but below 10<sup>-3</sup> M  $[Cu^{II}G_4]_T$  the O<sub>2</sub> uptake rate is lessened. Figures 4 and 5 show the dependence of the initial rate upon  $Cu^{III}(H_{-3}G_4)^{-1}$ concentration and upon O2 concentration, respectively. In both plots there is a trend from first order toward zero order as the concentration of the reactant being varied rises. It is also clear from Figure 5 that the concentration at which the  $O_2$  dependence saturates increases with increasing concentration of  $Cu^{III}(H_{-3}G_4)^{-}$ .

Mechanism of  $Cu^{III}(H_{-3}G_4)^-$  Decomposition and of  $O_2$ Uptake. Electrochemically prepared  $Cu^{III}(H_{-3}G_4)^-$  takes up  $O_2$  as it decomposes at high pH. The concentration depen-



Figure 5. Dependence of the initial rate of  $O_2$  uptake on the  $O_2$  concentration. Solid lines calculated from eq 7 with  $k_1 = 3.6 \times 10^{-4}$  s<sup>-1</sup> and  $k_3/k_2 = 0.5$ . Conditions: pH 7.5 in 0.018 M lutidine buffer, 0.10 M NaClO<sub>4</sub>, 25.0 °C, 2.5 × 10<sup>-3</sup> M [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub>. [Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-1</sup>]<sub>i</sub>, M: 0, 0.5 × 10<sup>-3</sup>;  $\blacktriangle$ , 1.25 × 10<sup>-3</sup>;  $\blacklozenge$ , 2.5 × 10<sup>-3</sup>;  $\bigstar$ , 5.0 × 10<sup>-3</sup>.

dences of  $O_2$  uptake in Figures 4 and 5 suggest a similar process. Equations 4, 5, and 6 can explain the kinetics where

$$\operatorname{Cu}^{\operatorname{III}}(\operatorname{H}_{-3}\operatorname{G}_4)^- \xrightarrow{k_1} \operatorname{R} + \operatorname{H}^+$$
(4)

$$R + O_2 \xrightarrow{\kappa_2} RO_2$$
 (5)

$$R + Cu^{III}(H_{-3}G_4)^{-} \xrightarrow{\kappa_3} Cu^{II}(H_{-1}G_4DHP) + Cu^{II}(H_{-3}G_4)^{2-} (6)$$

*R* is a reactive intermediate (either a carbon-centered free radical or a Cu(I) complex),  $RO_2$  is either a peroxy radical or a copper(III) peroxide, and G<sub>4</sub>DHP refers to the dehydropeptide given in eq 1. Treatment of R as a steady-state intermediate gives eq 7.

$$\frac{d[O_2]}{dt} = \frac{k_1 k_2 [Cu^{III}(H_{-3}G_4)^-][O_2]}{k_2 [O_2] + k_3 [Cu^{III}(H_{-3}G_4)^-]}$$
(7)

This rate expression is in qualitative agreement with the initial rate data in Figures 4 and 5. As a test of the quantitative agreement, these data were fit to eq 7 by a nonlinear regression analysis, which gives  $k_1 = 3.6 \times 10^{-4} \, \text{s}^{-1}$  and  $k_3/k_2 = 0.5$ . The quality of the fit was reasonable as may be seen from the solid curves in Figures 4 and 5.

Equations 4 and 5 represent the  $O_2$  uptake process while eq 4 and 6 represent the decomposition of  $Cu^{III}(H_{-3}G_4)^-$ . If  $k_1 << k_2[O_2]$  and if  $k_1 << k_3[Cu^{III}(H_{-3}G_4)^-]$ , then the rates of  $Cu^{III}(H_{-3}G_4)^-$  decomposition and of  $O_2$  uptake will be related. Decomposition of  $Cu^{III}(H_{-3}G_4)^-$  will have a rate constant equal to  $2k_1$  in the absence of  $O_2$  while the rate of  $O_2$  uptake is given by eq 7.

A further quantitative test of the agreement between experiment and the proposed mechanism is obtained by comparing values of  $k_1$  measured by Cu<sup>III</sup> decomposition to those calculated from initial rates of O<sub>2</sub> uptake using eq 7 and the value of  $k_3/k_2$  obtained from the nonlinear fit. As seen in Figure 6 there is agreement between the values of  $k_1$  obtained from O<sub>2</sub> uptake and from Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>) decomposition at different initial [Cu<sup>IIG</sup>G<sub>4</sub>]<sub>T</sub> concentrations. Both processes have



**Figure 6.** Values of  $k_1$  as a function of  $[Cu^{II}G_4]_T$  obtained from the initial rate of decomposition of  $Cu^{III}(H_{-3}G_4)^-$  (O) and from O<sub>2</sub> uptake ( $\Delta$ ). Conditions: pH 7.5 in 0.018 M 2,6-lutidine buffer, 0.10 M NaClO<sub>4</sub>, 25.0 °C. Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> data under N<sub>2</sub>. O<sub>2</sub> uptake data with  $1.2 \times 10^{-3}$  M O<sub>2</sub>.

Table IV. Determination of the Rate Constant for  $Cu^{III}(H_{-3}G_4)^-$  Decomposition at 25.0 °C and 0.1 M NaClO<sub>4</sub>

evaluated from	10 <sup>4</sup> k <sub>1</sub> , s <sup>-1</sup>	$[Cu^{II}G_4]_{T}, M$	buffer medium
initial rate of O, uptake	3.6	$2.5 \times 10^{-3}$	2,6-lutidine, pH 7.5
initial rate of Cu <sup>III</sup> decompn	3.5	$(2.5-5.0) \times 10^{-3}$	2,6-lutidine, pH 7.5
autocatalytic Cu <sup>III</sup> formn	2.8	$2.0 \times 10^{-3}$	unbuffered, pH 8.0
autocatalytic Cu <sup>III</sup> formn	2.8	$5.0 \times 10^{-4}$	unbuffered, pH 7.0
autocatalytic Cu <sup>III</sup> formn	2.8	8.0 × 10 <sup>-4</sup>	borate, pH 8.6
first-order Cu <sup>III</sup> decompn	2.4	$8.0 \times 10^{-4}$	borate, pH 8.6

approximately the same limiting rate at high  $[Cu^{II}G_4]_T$ concentrations and the same dependence upon  $[Cu^{II}G_4]_T$ concentrations. These data show that the  $k_1$  value is dependent on the  $[Cu^{II}G_4]_T$  concentration.

**Proposed Mechanism of Cu<sup>III</sup>(H**<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> Formation. The Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> decomposition and O<sub>2</sub> uptake data are consistent with the steps given in eq 4–6. The autocatalytic formation of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> requires further explanation. The reactive intermediate RO<sub>2</sub> is a strong oxidizing agent which can generate additional Cu(III) as well as the oxidized ligand (dehydropeptide) as shown in eq 8. If  $k_2[O_2] >> k_3$ -

$$RO_{2} + 3Cu^{II}(H_{-3}G_{4})^{2-} \xrightarrow{\text{rapid steps}} Cu^{II}(H_{-1}G_{4}DHP) + 3Cu^{III}(H_{-3}G_{4})^{-} (8)$$

 $[Cu^{III}(H_{-3}G_4)^-]$ , then the contribution from eq 6 will be small and the sum of eq 4, 5, 7, and 8 gives eq 9 which accounts for the autocatalytic production of  $Cu^{III}(H_{-3}G_4)^-$ . Equation 9

$$Cu^{III}(H_{-3}G_4)^- + 3Cu^{II}(H_{-3}G_4)^2 + O_2 + 4H^+ \rightarrow 3Cu^{III}(H_{-3}G_4)^- + Cu^{II}(H_{-1}G_4DHP) + 2H_2O (9)$$

shows that in the presence of  $O_2$  the decomposition of one  $Cu^{III}(H_{-3}G_4)^-$  results in a net gain of two  $Cu^{III}(H_{-3}G_4)^-$ . This accounts for the autocatalytic plots such as Figure 3. If eq 4 is the rate-determining step, the entire process is first order in  $Cu^{III}(H_{-3}G_4)^-$  and independent of the concentration of other reactants in accord with eq 3.

Table IV summarizes values of  $k_1$  determined by three distinct methods: (1) O<sub>2</sub> uptake rates, (2) Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> decomposition rates in the absence of O<sub>2</sub>, and (3) Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> autocatalytic formation in the presence of O<sub>2</sub>. Data



obtained under the same conditions (pH,  $[Cu^{II}G_4]_T$ , and the buffer used) give the same  $k_1$  values.

**Copper(II) Tetraglycine Catalysis.** The autocatalytic Cu<sup>III</sup> formation exhibits a  $[Cu^{II}G_4]_T$  dependence which saturates at high  $[Cu^{II}G_4]_T$  concentrations (Figure 2 and Table III) as previously seen for O<sub>2</sub> uptake and Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> decomposition rate constants in Figure 6. Both first-order Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> decomposition and autocatalytic Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> formation exhibit a catalysis by Cu(II) which saturates at 2 × 10<sup>-4</sup> M  $[Cu^{II}G_4]_T$  (Figure 2). The limiting rates for both processes are nearly equal; however, the agreement is not perfect.

The major Cu(II) complex over the pH range for the autoxidation is  $Cu^{II}(H_{-2}G_4)^-$ . It is proposed that the catalysis by Cu(II) involves the oxidation of this species by Cu<sup>III</sup>( $H_{-3}G_4$ )<sup>-</sup> (eq 10). As the structures in Scheme I indicate,

$$Cu^{II}(H_{-2}G_4)^- + Cu^{III}(H_{-3}G_4)^- \xrightarrow{k_4}_{k_{-4}}$$
  
 $Cu^{III}(H_{-2}G_4) + Cu^{II}(H_{-3}G_4)^{2-}$  (10)

eq 10 is not a proton-transfer reaction (which would require substitution reactions with sluggish Cu(III) complexes) but is an uphill electron-transfer reaction (on the basis of the variation of  $E^{\circ}$  values with ligand donors,<sup>5</sup> eq 10 would have a  $\Delta E^{\circ}$  value of about -0.3 V). Other reactions of Cu<sup>III</sup> peptides have very rapid electron-transfer rate constants<sup>6</sup> and this pathway could be important if Cu<sup>III</sup> (H<sub>-2</sub>G<sub>4</sub>) decomposes rapidly (eq 11). The higher oxidizing power of the Cu<sup>III</sup>-

$$Cu^{III}(H_{-2}G_4) + OH^- \xrightarrow{k_5} R + H_2O$$
(11)

 $(H_{-2}G_4)$  species compared to  $Cu^{III}(H_{-3}G_4)^-$  should favor more rapid ligand oxidation. A similar effect is observed in the greater instability of copper(III) tripeptide complexes compared to copper(III) tetrapeptide complexes.<sup>5</sup> Treatment of  $Cu^{III}(H_{-2}G_4)$  as a steady-state intermediate in eq 10 and 11 gives the expanded expression for  $k_1$  in eq 12, where  $k_1^\circ$  is

$$k_{1} = k_{1}^{\circ} + \frac{k_{4}k_{5}[OH^{-}][Cu^{II}(H_{-2}G_{4})^{-}]}{k_{-4}[Cu^{II}(H_{-3}G_{4})^{2^{-}}] + k_{5}[OH^{-}]}$$
(12)

the value of  $k_1$  as the concentration of  $\operatorname{Cu}^{II}(\operatorname{H}_2G_4)^-$  approaches zero (i.e., a Cu(II)-independent pathway as shown in the intercept of Figure 2). Letting  $[\operatorname{Cu}^{II}(\operatorname{H}_3G_4)^{2-}] = (K_3/K_w)[\operatorname{Cu}^{II}(\operatorname{H}_2G_4)^-][\operatorname{OH}^-]$ , where  $K_3$  is the ionization constant for the first peptide nitrogen, and substituting into eq 12 gives

$$k_{1} = k_{1}^{\circ} + \frac{k_{4}k_{5}[\operatorname{Cu}^{II}(\operatorname{H}_{-2}\operatorname{G}_{4})^{-}]}{(K_{3}/K_{w})k_{-4}[\operatorname{Cu}^{II}(\operatorname{H}_{-2}\operatorname{G}_{4})^{-}] + k_{5}}$$
(13)

This corresponds to the observed behavior in which the  $k_1$  value is pH independent and the copper(II) catalysis saturates at

## Autoxidation of Copper(II) Tetraglycine





Scheme III



high concentrations  $(k_1 = k_1^{\circ} + (k_4/k_{-4})k_5)$ . From the data in Table III the ratio  $k_5/k_{-4} \approx 4$ . The fact that the ligand oxidation occurs predominantly at the third glycyl residue (rather than at the fourth glycyl residue as is the case with nickel) is consistent with the decomposition of Cu<sup>III</sup>(H<sub>-2</sub>G<sub>4</sub>), in which only three nitrogens (an amine group and two deprotonated peptides) are coordinated. Equation 11 corresponds to the loss of a methylene proton from the third glycyl residue and the reduction of the Cu(III) center as shown in Schemes II and III.

**Buffer Medium Effects.** The kinetics of  $Cu^{III}(H_{-3}G_4)^$ formation and decomposition are dependent upon the buffer medium. Data for borate buffer are comparable to those for unbuffered solutions. By comparison, in 2,6-lutidine buffer at pH 7.5 the rates of O<sub>2</sub> uptake and of Cu(III) decomposition (Figure 6) require higher [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub> concentrations to attain the limiting values. This effect is much more pronounced in Tris buffer. The rate of decomposition of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> at low [Cu<sup>II</sup>G<sub>4</sub>]<sub>T</sub> concentrations is diminished. The autoxidation reaction does not occur at all in 0.010 M Tris.

An Additional  $O_2$  Dependence. At lower  $O_2$  concentrations the data in Table III show an  $O_2$  dependence even when  $k_2[O_2]$ >>  $k_3[Cu^{III}(H_{-3}G_4)^-]$ . Hence this  $O_2$  dependence cannot be due to the competition between the rates in eq 5 and 6. A parallel path in competition with eq 5 is needed. This competitive path cannot be the reverse of eq 4 since this would cause the decomposition of  $Cu^{III}(H_{-3}G_4)^-$  to deviate from a first-order dependence. A reaction is proposed (eq 14) in which

$$\mathbf{R} \xrightarrow{\kappa_6} \mathbf{P} \tag{14}$$

R decays to a product P which is less reactive with  $O_2$  but may still react rapidly with Cu(III). The nature of the species R,  $RO_2$ , and P depends upon the choice between a radical intermediate pathway and a copper(I) dehydropeptide pathway.

Radical Intermediate Pathway. The ionization of a methylene proton from the third glycyl residue of a Cu(III) complex could produce a Cu(II) complex with a carboncentered radical as shown in Scheme II. This radical (R) could react with  $O_2$  ( $k_2$  step) to give a peroxy radical (RO<sub>2</sub>) which would decay in a series of steps to give more  $Cu^{III}(\tilde{H}_{-3}G_4)^$ and the dehydropeptide complex,  $Cu^{II}(H_{-1}G_4DHP)$ . Higher concentrations of  $\hat{C}u^{III}(H_{-3}G_4)^-$  would convert the radical to the dehydropeptide  $(k_3 \text{ step})$  in competition with the O<sub>2</sub> reaction. Low concentrations of both  $O_2$  and  $Cu^{III}(H_{-3}G_4)^-$  could cause an internal conversion ( $k_6$  step) to the copper(I) dehydropeptide complex. The RO<sub>2</sub> complex undergoes a series of electron transfers and decomposition steps to regenerate  $Cu^{III}(H_{-3}G_4)^-$  and to give the copper(II) dehydropeptide complex. In this mechanism the radical R must be sufficiently long-lived to have the  $O_2$  and Cu(III) steps compete with the rate of internal electron transfer to give the copper(I) dehydropeptide.

The data in Table III indicate that  $k_6 \approx k_2[O_2]$  when the  $O_2$  concentration is  $10^{-4}$  M. Since  $k_2$  cannot be larger than  $7 \times 10^9$  M<sup>-1</sup> s<sup>-1</sup> (the diffusion-controlled rate constant), this sets a maximum possible value of  $7 \times 10^5$  s<sup>-1</sup> for  $k_6$ . Rate constants for internal electron transfer between Co(III) and other metal centers in binuclear complexes<sup>30</sup> can be many orders of magnitude smaller than this. However, the  $k_6$  rate constant in Scheme II is for the transfer of an electron from a high-energy free-radical species to a nearby copper(II). This process might be expected to be very fast, and the limiting value for  $k_6$  appears to be one difficulty with the radical intermediate mechanism.

**Copper(I) and Copper(III) Peroxide Intermediate Pathway.** Scheme III gives an alternate mechanism based on a very short-lived Cu<sup>II</sup>-radical species which decays to a copper(I) dehydropeptide species as a direct result of the  $k_1$  step. This could be considered as an internal two-electron transfer. In this mechanism the chain center R is Cu<sup>I</sup>(H<sub>-1</sub>G<sub>4</sub>DHP) and the Cu(I) complex must react rapidly with O<sub>2</sub> to give a copper(III) peroxide complex as the RO<sub>2</sub> intermediate. This mechanism as well as that in Scheme II is subject to catalysis by Cu<sup>II</sup>(H<sub>-2</sub>G<sub>4</sub>)<sup>-</sup>. The resulting rate expressions are the same, but in Scheme III the reaction center is at copper rather than at carbon.

Most rate constants for the reaction of  $O_2$  with Cu(I) complexes are in the range of  $10^2-10^4 \text{ M}^{-1} \text{ s}^{-1}.^{31}$  However, these constants are for reactions believed to be one-electron transfers to give Cu<sup>II</sup>(O<sub>2</sub><sup>-</sup>) rather than the proposed two-electron transfer to give Cu<sup>III</sup>(O<sub>2</sub><sup>2-</sup>). The  $k_2$  rate constant must be nearly the same as the  $k_3$  rate constant to fit the observed behavior. Rate constants for the reaction of bis(2,9-dimethyl-1,10-phenanthroline)copper(I) with copper(III)-peptide complexes are as large as  $10^6 \text{ M}^{-1} \text{ s}^{-1}.^{32}$  If  $k_3$  is this large, then  $k_2$  must also be large and this requirement may be a difficulty with this mechanism. On the other hand, the  $k_6$  step is a dissociation process (or perhaps a partial unwrapping of the peptide from Cu(I)) which could be slow enough to compete with the  $k_2$  and  $k_3$  steps. Hence there are pros and cons for both mechanisms.

**Initiation Reaction.** The proposed mechanisms for the autoxidation provide a rationale for the production of significant amounts of  $Cu^{III}(H_{-3}G_4)^-$  from an initially very small

 $(10^{-7}-10^{-8} \text{ M})$  concentration of Cu<sup>III</sup> or other oxidizing agent.

In the initiation experiments, solutions of  $[Cu^{II}G_4]_T$  were swept with  $N_2$  and stoppered overnight. Since the Cu<sup>II</sup>G<sub>4</sub>catalyzed decomposition of  $Cu^{III}(H_{-3}G_4)^-$  has  $k_{obsd} = 2k_1 (\approx 5.0$  $\times$  10<sup>-4</sup> s<sup>-1</sup>), any small quantities of Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup> formed during solution preparation from oxidizing impurities would be completely decomposed before the solution was used. This excludes initiation by oxidizing impurities.

The direct reaction of  $O_2$  with  $Cu^{II}(H_{-3}G_4)^{2-}$  to give  $Cu^{III}(H_{-3}G_4)^-$  and  $O_2^-$  is an unlikely initial step because the equilibrium levels of  $Cu^{III}$  and  $O_2^-$  would be only  $10^{-13}$  M. This can be estimated from the electrode potential for the reduction of  $O_2^{33}$  and of  $Cu^{III}(H_{-3}G_4)^{-5}$ . It is possible for two- or four-electron reactions with  $O_2$  to provide sufficient Cu<sup>III</sup>, but if these reactions could occur rapidly then there would be no need for the autocatalytic pathway which is observed.

Furthermore, the DPPH experiments showed that  $O_2$  did not affect the difference in induction times caused by the addition of this radical scavenger. Hence, it appears that the initiation reaction does not involve  $O_2$ .

We propose that the initiation mechanism involves the disproportionation of Cu(II) to give Cu(I) and Cu(III). A very interesting paper by Österberg<sup>34</sup> has shown that the copper triglycine system forms a mixed Cu(I) and Cu(II) complex with a favorable reduction potential ( $E^{\circ} = +0.34$  V at pH 7 for eq 15). If  $G_4$  reacted in the same manner as  $G_3$  then the

$$Cu_2^{II}H_{-4}(G_3)_2^{2-} + H^+ + e^- → Cu^1Cu^{II}H_{-3}(G_3)_2^{2-}$$
 (15)

half-reaction in eq 16  $(E^{\circ} = -0.63 \text{ V})^5$  could be coupled with

$$Cu^{II}(H_{-3}G_4)^2 \rightarrow Cu^{III}(H_{-3}G_4)^- + e^-$$
 (16)

a reaction of the type given in eq 15. Writing these reactions in terms of a  $Cu^{II}(H_{-2}G_4)^{2-}$  monomer leads to eq 17 which  $3Cu^{II}(H_{-2}G_4)^- \rightleftharpoons Cu^{IC}u^{II}H_{-3}(G_4)_2^{2-} + Cu^{III}(H_{-3}G_4)^-$  (17)

would have an equilibrium constant of 10<sup>-6.8</sup> M<sup>-1</sup>. This predicts that a concentration of  $10^{-3}$  M Cu<sup>II</sup>(H<sub>-2</sub>G<sub>4</sub>)<sup>2-</sup> would produce  $1.3 \times 10^{-8}$  M Cu<sup>III</sup>(H<sub>-3</sub>G<sub>4</sub>)<sup>-</sup>, which is not far from the experimental value of 10<sup>-7</sup> M Cu(III). However, a log-log plot of the initial Cu(III) found (varying from  $10^{-8}$  to  $10^{-6.5}$  M as measured from the intercepts of the autocatalytic plots) vs. the concentration of  $[Cu^{II}\hat{G}_4]_T$  (from 10<sup>-4</sup> to 10<sup>-2.5</sup> M) gives a slope of unity in accord with eq 18 rather than a slope of

$$2Cu^{II}(H_{-2}G_4)^- \rightleftharpoons Cu^{I}(H_{-1}G_4)^- + Cu^{III}(H_{-3}G_4)^-$$
 (18)

 $^{3}/_{2}$  expected from eq 17. The concentrations used in the present study are much lower than those used by Österberg and therefore monomeric species are more likely. The identity of the Cu(I) complex is uncertain and might also be  $Cu^{I}G_{4}(OH)$  or even  $Cu^{I}(OH)$ . An equilibrium constant 10<sup>-8</sup> for eq 18 fits our data. The fact that Cu(III) persists in trace quantities no matter how carefully  $[Cu^{II}G_4]_T$  solutions are prepared or how long they are kept under N2 strongly supports initiation by a disproportionation mechanism.

Comparison of  $Cu^{II}G_4$  and  $Ni^{II}G_4$  Autoxidations. The mechanisms for autoxidation for both the  $Cu^{II}G_4$  and  $Ni^{II}G_4$ reactions are dependent on the decomposition of the trivalent metal complex. The rate of decomposition of  $Ni^{III}(H_{-3}G_4)^$ is approximately 20 times greater than that of  $Cu^{III}(H_{-3}G_4)^$ and this accounts for the faster autoxidation of  $Ni^{II}G_4$ . The  $Ni^{III,II}(H_{-3}G_4)$  potential of 0.79 V is higher than the  $Cu^{III,II}(H_{-3}G_4)$  potential of 0.63 V. This may be related to the fact that decarboxylation occurs in the nickel system but not in the copper system. The need for a higher potential to generate the reactive intermediate (a carbon-centered free radical or a Cu(I) species) is consistent with the hypothesis that the reactive copper species is  $Cu^{III}(H_{-2}G_4)$  formed by uphill electron transfer.

## Conclusions

The overall autoxidation rate expression (except at low concentrations of  $O_2$  and of  $Cu^{II}(H_2G_4)^{-}$  is given in eq 19.

$$-d[O_{2}]/dt = k_{2}k_{4}k_{5}[Cu^{III}(H_{-3}G_{4})^{-}][Cu^{II}(H_{-2}G_{4})^{-}][O_{2}]/ \{(K_{3}/K_{w})k_{-4}[Cu^{II}(H_{-2}G_{4})^{-}] + k_{5}\}\{k_{2}[O_{2}] + k_{3}[Cu^{III}(H_{-3}G_{4})^{-}]\} (19)$$

The  $Cu^{III}(H_{-3}G_4)^-$  appears to be generated at trace levels by the disproportionation of  $Cu^{II}(H_{-2}G_4)^-$  not shown in eq 19. The  $Cu^{II}(H_{-2}G_4)^-$  complex also catalyzes the Cu(III) decomposition process which is the rate-determining step for the entire autoxidation. This catalysis requires rapid electrontransfer reactions. The multiple oxidation states of copper, the ability to use copper(III) peptides as a reservoir of oxidizing power, and the rapid electron-exchange reactions of copper make it an effective catalyst in this autoxidation.

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Registry No. Cu<sup>II</sup>(H<sub>2</sub>G<sub>4</sub>)<sup>-</sup>, 67180-35-2; Cu<sup>III</sup>(H<sub>3</sub>G<sub>4</sub>)<sup>-</sup>, 57692-61-2.

### **References and Notes**

- (1) Kaneda, A.; Martell, A. E. J. Coord. Chem. 1975, 4, 137.
- Freeman, H. C.; Taylor, M. R. Acta Crystallogr. 1965, 18, 939. Youngblood, M. P.; Bannister, C. E.; Chellappa, K. L.; Margerum, D. (2)
- (3) W., to be submitted for publication.
- (4) Margerum, D. W.; Chellappa, K. L.; Bossu, F. P.; Burce, G. L. J. Am. Chem. Soc. 1975, 97, 6894
- (5) Bossu, F. P.; Chellappa, K. L.; Margerum, D. W. J. Am. Chem. Soc. 1977, 99, 2195
- (6) Owens, G. D.; Chellappa, K. L.; Margerum, D. W., to be submitted for publication.
- (7) Burce, G. L.; Paniago, E. B.; Margerum, D. W. J. Chem. Soc., Chem. Commun. 1975, 261
- Paniago, E. B.; Weatherburn, D. C.; Margerum, D. W. Chem. Commun. (8) 1971, 1428.
- (9) Bossu, F. P.; Paniago, E. B.; Margerum, D. W.; Kirksey, S. T., Jr.; Kurtz, Dossi, F. F., Fainago, E. B., Margeruni, D. W., Kirksey, S. L., Jr., Kur J. L. Inorg. Chem. 1978, 17, 1034.
   Gillard, R. D.; Spencer, A. Discuss. Faraday Soc. 1968, 46, 213.
   Gillard, R. D.; Spencer, A. J. Chem. Soc. A 1969, 2718.
   Gillard, R. D.; Phipps, D. A. J. Chem. Soc. A 1971, 1074.
- (10)
- (11)
- (12)(13)
- Michaelidis, M. S.; Martin, R. B. J. Am. Chem. Soc. 1969, 91, 4683. McKenzie, E. D. J. Chem. Soc. A 1969, 1655. (14)
- (15)Caglioti, V.; Silvestioni, P.; Furlani, C. J. Inorg. Nucl. Chem. 1960, 13,
- (16) Harris, W. R.; Bess, R. C.; Martell, A. E.; Ridgeway, T. H. J. Am. Chem. (10) Harris, W. R.; Bess, R. C., Hartell, A. E., Ridgeway, T. H. J. Am. Chem. Soc. 1977, 99, 2958.
  (17) Harris, W. R.; Martell, A. E. J. Am. Chem. Soc. 1977, 99, 6746.
  (18) Beck, M. T.; Görög, S. Acta Chim. Acad. Sci. Hung. 1961, 21, 401.
  (19) Lock, M. V.; Sagar, B. F. J. Chem. Soc. B 1966, 690.
  (20) Sagar, B. F. J. Chem. Soc. B 1967, 428.

- Sagar, B. F. J. Chem. Soc. B 1967, 1047.
   Sagar, C. L.; Jancous, J. J.; Jayasinhuler, K. Aust. J. Chem. 1972, 25, 1819
- (23) Milner, O. I. Analysis of Petroleum for Trace Elements; Macmillan: New York, N.Y., 1963; p 50.
- Hamilton, G. A. Adv. Enzymol. Relat. Subj. Biochem. 1969, 32, 55.
- (25) Bossu, F. P.; Margerum, D. W. Inorg. Chem. 1977, 16, 1210.
- Anbar, M.; Levitzki, A.; Berger, A. Biochemistry 1967, 6, 3757.
- (27) Kurtz, J. L.; Rybka, J. S.; Neubecker, T. A.; Margerum, D. W., to be (2) Karte, J. A. Ryski, V. J. Kolsekker, F. H., Malgerein, D. W. & Cossubmitted for publication.
  (28) Guilbaut, G., Kramer, D. N.; Hackley, E. Anal. Chem. 1967, 39, 271.
  (29) Levitzki, A.; Berger, A. Biochemistry 1971, 10, 64.
  (30) Haim, A. Acc. Chem. Res., 1975, 8, 264.
  (31) Crumbliss, A. L.; Gestant, L. J. J. Coord. Chem. 1976, 5, 109.

- (32) Lappin, A. G.; Youngblood, M. P.; Margerum, D. W., to be submitted for publication.
- Wood, P. M. FEBS Lett. 1974, 44, 22. (33)
- (34) Österberg, R. Eur. J. Biochem. 1970, 13, 493.